

ISSN 1477-9315



JOURNAL OF

ENVIRONMENTAL HEALTH RESEARCH

Journal of environmental health research. Volume 1 Issue 3 2022

ISSN 1477-9315 <http://www.jehr-online.org/>

<https://doi.org/10.5281/zenodo.7408374>

[Universal impact factor 7.2](#)

Journal of environmental health research. ISSN 1477-9315

The abbreviation of the journal title "**Journal of environmental health research**" is "**J. Environ. Health Res.**". It is the recommended abbreviation to be used for abstracting, indexing and referencing purposes and meets all criteria of the [ISO 4 standard](#) for abbreviating names of scientific journals.

Journal of Environmental Health Research is devoted to the rapid publication of research in environmental health, acting as a link between the diverse research communities and practitioners in environmental health. Published articles encompass original research papers, technical notes and review articles. JEHR publishes articles on all aspects of the interaction between the environment and human health. This interaction can broadly be divided into three areas: 1. The natural environment and health – health implications and monitoring of air, water and soil pollutants and pollution and health improvements and air, water and soil quality standards; 2. The built environment and health – occupational health and safety, exposure limits, monitoring and control of pollutants in the workplace, and standards of health; and 3. Communicable diseases – disease spread, control and prevention, food hygiene and control, and health aspects of rodents and insects.

Editorial board

Professor Chan Lu – Xiang Ya School of Public Health, Central South University, China
Dr. Kristina Mena - School of Public Health, the University of Texas Health Science Center at Houston, USA
Dr Pablo Orellano - National Scientific and Technical Research Council (CONICET) and National Technological University, Argentina
Abdumalik Djalilov Tashkent Pediatric Medical Institute
Dilfuza Turdieva Tashkent Pediatric Medical Institute
Nigora Alieva Tashkent Pediatric Medical Institute
Khursandoy Akramova Tashkent Pediatric Medical Institute
Ozimbay Otaxanovich Jabbarov Tashkent medical academy
Professor Susan Pinney – College of Medicine, University of Cincinnati, USA
Professor Grażyna Plaza – Institute for Ecology of Industrial Areas, Poland
Professor Andrew Povey – School of Health Sciences, University of Manchester, UK
Dr Jack Siemiatycki - University of Montreal, Canada
Dr. Baltabaev Ubaidulla Abduvakilovich Tashkent State Dental Institute
Dr. Asrankulova Diloram Bakhtiyarovna - doctor of medical sciences, associate professor. Andijan State Medical institute
Dr. KHudaynazarova Salomat Tashkent Pediatric Medical Institute, Hospital Pediatrics 2, Department of Folk Medicine. PhD
Dr. Rakhimov Oybek Umarovich Tashkent Pediatric Medical Institute
Dr. Jafarov Khasan Mirzakhidovich, Tashkent Pediatric Medical Institute
Dr. Sodikova Dilrabo Andijan state medical institute
Dr. Kutlikova Gusalhon Andijan state medical institute
DSc, Musashaykhov Khusanboy Tadjibaevich Andijan State Medical Institute
Raimkulova Narina Robertovna Tashkent Pediatric Medical Institute
Nasirova Feruza Jumabaevna Andijan State Medical Institute

Manuscripts typed on our article template can be submitted through our website here. Alternatively, authors can send papers as an email attachment to editor@jehr-online.org

Journal of environmental health research.
ISSN 1477-9315 <http://www.jehr-online.org/>
36 Victoria Road London N59 7LB

The state of the antimicrobial system of the oral cavity in patients with insulin resistance

Ziyatova G.Z, Rustamova S.M, Khajimetov A.A, Atakhadjayeva M.A,
Akhmadaliyev N.N.

(Tashkent State Dental Institute)

Ziyatova Guzal Zarifbayevna - Supporting doctoral student of the department of
medical and biological chemistry, TSDI.

Rustamova Sabogul Mamarejabovna - Supporting doctoral student of the
department of medical and biological chemistry, TSDI.

Khajimetov Abdugafur Akhmatovich - Doctor of biology, professor, Tashkent
State Dental Institute

Atakhadjayeva M.A- Candidate of chemical sciences, Tashkent State Dental
Institute

Akhmadaliyev Nusrat Nugmanovich - Head of the department of medical and
biological chemistry, TSDI.

ABSTRACT

The aim of this study was to evaluate the antimicrobial activity of oral fluid peptides in patients with insulin resistance. The objects of the study were 32 patients who were divided into the main study group and the control group. Of these, 18 patients were included in the main group at the stages of treatment for insulin resistance, in which a high level of C-peptide in the blood was detected by enzyme immunoassay. In the oral fluid, the amount of the antimicrobial peptide, cathelicidin LL-37, alpha-defensin 1-3, was assessed by enzyme immunoassay. In studies in the oral fluid of patients with IR, instead of a pronounced increase in the antimicrobial peptide, a decrease in its concentration was revealed, which requires additional studies.

Introduction

At present, the actual problem is the knowledge of the oral fluid as a biologically significant microenvironment of the body from the standpoint of a multidisciplinary approach, its metabolic and immunological profile. Taking into account the availability of oral fluid, the non-invasiveness of obtaining it, the possibility of multiple dynamic research, this bioenvironment is increasingly of interest as an object of study in fundamental and clinical practice as an alternative to blood in the diagnosis of many diseases.

It should be noted that the arguments for choosing the oral fluid as an object for studying the body are the following: being a complex filtrate of blood plasma, it reflects the state of dynamic constancy of the internal environment of the body; is an indicator of the reactivity of the organism, as it can change in composition, physico-chemical and biological properties under the influence of various factors.

As is known, blood components enter the oral cavity through the fluid of the periodontal sulcus, mucous and mucous transudate and through bleeding that occurs in the oral cavity. As a result, a huge molecular diversity is found in the oral cavity, often referred to as "mixed saliva". Mixed saliva plays an important role for both physicochemical and immune protection of the oral mucosa through direct antimicrobial activity and agglutination of microorganisms. At the same time, saliva proteins are multifunctional in nature to protect the oral mucosa, as well as soft and hard tissues of the oral cavity. Antimicrobial peptides (AMPs) occupy a special place among the protective factors of the oral cavity. These are small molecules containing from 12 to 50 amino acid residues that can kill microbial cells. Most currently known AMPs. These are small molecules containing from 12 to 50 amino acid residues that can kill microbial cells. Most currently known AMPs have a broad spectrum of antimicrobial activity, acting against gram-positive and gram-negative bacteria, as well as yeasts and some viruses. In addition, convincing evidence has been obtained that a number of AMPs have anticarcinogenic activity and are also immunomodulators. Mixed saliva proteins provide antimicrobial protection by binding to bacteria and lysing microbial walls. They may also be responsible for the antifungal and antiviral properties of saliva. It has now become clear that AMPs in the oral cavity not only destroy pathogenic microorganisms, but also participate in the maintenance of normal microflora.

Currently, the following types of AMPs have been found in the oral cavity: α - and p-defensins, histatins, adrenomedullin and human cathelicidins, the sources of which are the oral mucosa, salivary glands and neutrophils protein calprotectin complement the protective function of antimicrobial factors of the salivary glands, lysozyme, immunoglobulins and histatins.

It should be noted that the microflora of the human oral cavity is extremely diverse and is normally represented by several hundred species of microorganisms. In periodontal diseases, as a rule, the quantitative ratio of microbes changes, and their species composition remains constant. This circumstance suggests that the cause of diseases of the tissues of the oral cavity is not actually a bacterial infection, but a violation of the adequate interaction of the macroorganism with the microflora. In this regard, researchers are of particular interest in the study of the protective systems of the oral cavity, in particular, antimicrobial peptides of the oral fluid.

The aim of this study was to evaluate the antimicrobial activity of oral fluid peptides in patients with insulin resistance.

Material and research methods

The objects of the study were 32 patients who were divided into the main study group and the control group. Of these, group I included 18 patients (12 men and 6

women) aged 18 to 65 years (average age 47.6 years), who were at the stages of treatment for insulin resistance in whom the enzyme immunoassay revealed a high level of C-peptide in the blood, group 2 (14) consisted of healthy individuals aged 18 to 65 years (9 men and 5 women) without concomitant pathology. A comprehensive dental and laboratory study of patients in groups 1 and 2 was carried out at the clinical bases of TSDI and TMA.

To assess the state of insulin resistance, blood was taken in the morning, on an empty stomach. The content of insulin and the level of C-peptide in the blood serum were determined by enzyme immunoassay using firm from HUMAN. The concentration of insulin on an empty stomach was taken as the norm of C-peptide, which amounted to 0.78 -1.89 ng / ml (SI: 0.26-0.63 mmol / l). HOMA insulin resistance index (HOMA IR) = (fasting blood insulin x fasting blood glucose) / 22.5; (norm < 2.5). In our opinion, the determination of C-peptide is preferred in the diagnosis of insulin resistance in patients, since the concentration of insulin in the blood is about 5 times less than that of C-peptide, while the duration of the "life" of the insulin molecule is 4 minutes, C-peptide - 20 minutes.

The intake of oral fluid was carried out on an empty stomach from 8 to 9 hours and was carried out by spitting into a glass, sterile test tube for 5 minutes, without its preliminary stimulation. The volume of oral fluid averaged 7 ml. The samples were then purified by centrifugation at 10,000 × g for 5 min from cells, frozen and stored at -30 C. On the day of analysis, the samples were thawed and further purified of mucin by centrifugation at 3000 × g for 5 min. In the oral fluid, the amount of antimicrobial peptide - cathelicidin LL-37, alpha-defensin 1-3, was assessed by enzyme immunoassay using a set of reagents from «ООО БюХимМак» Russia. Statistical processing of the material was carried out using standard software packages (statistics 6 0, Excel 2003). To determine the statistical significance of differences in continuous values depending on the distribution parameters Student's t-tests or the Mann-Whitney test were used. Differences were considered significant for all analyzes at a significance level of p<0.05.

Results and its discussion

Our data indicate that patients with IR have characteristic basal hyperinsulinemia, i.e. decreased sensitivity of peripheral tissues to insulin in the observation group. Apparently, the prescription of the disease and age affect the insulin receptors in peripheral tissues to the action of insulin. An indirect method for detecting IR also includes the determination of C-peptide on an empty stomach. The study of the level of C-peptide in the blood can serve as a more accurate confirmation of insulin hypersecretion in IR, since there are some known methodological limitations in the laboratory determination of insulin in the blood, which is 50% bound in the liver and has a half-life in the peripheral blood of about

4 minutes. The C-peptide is cleaved from the proinsulin molecule when it is converted into insulin, does not bind to cell receptors on the periphery, has a half-life of about 30 minutes, and is not extracted from the blood plasma by the liver. According to our studies, in the group of patients with IR, the level of C-peptide was significantly higher in relation to patients in the control group. Therefore, our results of the study indicate a direct relationship between the level of C-peptide in the blood of patients with IR.

Table 1**Criteria for insulin resistance and blood levels of adiponectin in examined patients**

Index	Patients with IR n=18	Healthy person n=14
Insulin (on an empty stomach) (mkED/l)	15,78±0,62*	12,61±0,47
C-peptide (ng/ml)	5,85±0,39*	1,51±0,16
HOMA index (kg/m ²)	4,01±0,25*	3,02±0,23
Adiponectin (mkg/ml)	4,12 ±0,41*	7,22±0,44

Notes: * - significance of differences $P < 0.05$

As you know, adipose tissue appears as a hormonally active organ, which is credited with the production of leptin, adiponectin, and resistin. Studies by numerous authors have shown that the secretion of adiponectin by adipocytes is stimulated by insulin. Adiponectin regulates energy homeostasis and has an anti-atherogenic and anti-inflammatory effect by inhibiting the adhesion of monocytes to vascular endothelial cells and exerting an inhibitory effect on growth factor-induced proliferation of smooth muscle cells in the vascular wall. As shown in our studies, the low content of adiponectin in the blood serum of patients of group 1 is an independent factor in the prognosis of the development of IR, since the expression of adiponectin does not correlate with blood C-peptide values.

Recently, scientists from the University of California, San Diego School of Medicine, USA, in a new study found that adipocytes, fat cells, are able to produce antimicrobial peptides that prevent the introduction of bacteria and pathogens of various origin. To confirm this version, we studied some antimicrobial peptides in patients with IR. At the same time, it was found that in persons with insulin resistance, the concentration of antimicrobial peptides in the blood was significantly lower relative to the comparison groups, which, in our opinion, is due to the low level of synthesis of complete antimicrobial peptides by adipocytes.

Recently, it has been proven that a-defensins have a unique spectrum of antimicrobial activity, in particular, they are highly effective against *Porphyromonas gingivalis*, which cause damage to periodontal tissues. Also, a-defensins have a pronounced antiviral activity against herpes, influenza, hepatitis C viruses, human immunodeficiency virus (HIV)-1, cytomegaloviruses, papillomaviruses, adenoviruses. They are present in the epithelium of the gums, tongue, salivary glands and mucous membranes. Modern literature reports that defensins directly affect the adhesion of microorganisms to periodontal tissues and oral mucosa, and hence the development of periodontitis and diseases of the oral mucosa.

As can be seen from the presented research results, the content of a-defensins 1-3 in the oral fluid in patients of the main group and in healthy individuals, presented in Table 2, indicates a reduced secretion of a-defensins 1-3 in the oral fluid of the main group relative to the comparison group. The revealed factual material indicates the inactivation of β -defensins, which can lead to an increase in microbial colonization and increases the risk of viral and bacterial infections in the oral cavity.

The antimicrobial peptide LL-37 has antibacterial properties and is basically expressed in neutrophil granules and epithelial cells. But at the same time, it can increase in biological media under stress, cell damage, contributing to wound healing and repair. The characteristics of the content of cathelicidin LL-37 in the oral fluid in patients of the main and control groups are presented in Table 2. The decrease in cathelicidin KL-37 in the oral fluid in patients of the main group can be explained by the following circumstance. As is known, cathelicidin LL-37 is synthesized in increased amounts in various lesions of the oral mucosa, which contributes to its faster healing. In our studies in the oral fluid, instead of a pronounced increase in the antimicrobial peptide, a decrease in its concentration in patients with IR was noted. The revealed factual material requires additional studies to explain the results of studies in patients with IR...

Table 2

Indicators of the content of antimicrobial peptides in the oral fluid in the examined patients with IR.

Indicators	Healthy person (n=14)	Patients with IR n=18
Cathelicidin LL-37 (ng/ml)	48,17±3,72	31,26±2,81*
Alpha defensins 1-3 (ng/ml)	984,2±±16,71	697,2±13,86*

Notes: * - significance of differences P<0.05

Thus, despite the fact that antimicrobial peptides, being a component of innate immunity, perform the function of the body's natural defense against a wide range of microbes in many human organs and systems, their multidirectional changes in patients with IR undoubtedly arouse interest in studying the pathogenetic mechanisms of their role. in the implementation of antimicrobial protection in IR. In addition, in our opinion, the study of antimicrobial peptides of oral fluid is the fact that peptides produced in response to bacterial invasion of the oral cavity can be identified and used as biomarkers for early diagnosis of the disease and its prevention.

Literature

1. Kazeko L.A. Possibilities of diagnosing periodontal diseases using antimicrobial peptides of saliva and gingival fluid // *Modern Dentistry*. - 2016. - №.1. - P.11-16.

2. Pantelev P. V., Bolosov I. A., Balandin S. V., Ovchinnikova T. V. Structure and biological functions of P-hairpin antimicrobial peptides. *Journal "ACTA NATURE"* -2015. - T. 7. - No. 1 (24). - P.37-47.

3. Ali Adem Bahar, Dacheng Ren. Antimicrobial Peptides//*Pharmaceuticals (MDPI)*. - 2013. - Vol.6 (12). - P.1543-1575.

4. Alexandre L. Pereira, Gilson C. Franco, Sheila C. Cortelli.. Influence of Periodontal Status and Periodontopathogens on Levels of Oral Human P-defensin-2 in Saliva // *Journal of Periodontology*.-2013. - V.84. - No. 1. - P.1445-1453.

5. Nagihan Bostanci, Georgios N. Belibasakis. Antimicrobial peptides. *Porphyromonas Gingivalis: An Invasive and Evasive Opportunistic Oral Pathogen*// *FEMS Microbiology Letters*. - 2012. - V.333. - No. 1. - P.1-9.

6. Bechinger B. Detergent-like Properties of Magainin Antibiotic Peptides: A 31P Solid-State NMR Spectroscopy Study// *Biochimica et Biophysica Acta*. - 2005. - V.1712. - No. 1. - P.101-108.

7. Bolintineanu D.S., Kaznessis Y.N. Computational studies of protegrin antimicrobial peptides: A review// *Peptides*. - 2011. - V.32. - No. 1. - P.188-201.

8. Brancatisano FL, Maisetta G, Barsotti F, Esin S, Miceli M, Gabriele M, Giuca MR, Campa M, Batoni G. Reduced Human Beta Defensin 3 in Individuals With Periodontal Disease// *Journal of Dental Research*. - 2011. - V.90. - No. 2. - P.241-245.

9. Gorr S.U., Kinane D.F., Mombelli A. Antimicrobial Peptides in Periodontal Innate Defense.// *Frontiers of oral biology*. - 2012. - V.15. - P.84-98.

10.Henrik Dommisch Soren Jepsen. Diverse Functions of Defensins and Other Antimicrobial Peptides in Periodontal Tissues // *Journal of Periodontology*. -2015.- V.69. - No. 1. -P.96-110.

11. Marie-Laure Jourdain, Frederic Velard. Cationic Antimicrobial Peptides and Periodontal Physiopathology: A Systematic Review // Journal of Periodontal Research.. -2019. - V.54. - No. 6. - P.589-600.

12. Martin Malmsten. Antimicrobial peptides// Upsala Journal of Medical Sciences. -2015.-V.119.-№2. -P.199-204.

13. S. Li G. Schmalz J. Schmidt F. Krause R. Haak D. Ziebolz Antimicrobial Peptides as a Possible Interlink Between Periodontal Diseases and Its Risk Factors: A Systematic Review//Journal of Periodontal Research. - 2018. - V.53. - No. 2. - P.145-155.

14. Xiaojing Xia Likun Cheng Shouping Zhang Lei Wang Jianhe Hu. The Role of Natural Antimicrobial Peptides During Infection and Chronic Inflammation // Journal Antonie van Leeuwenhoek. - 2018. - V.111. - No. 1. - P.5-26.

15. Hans M, Madaan Hans V. Epithelial antimicrobial peptides: guardian of the oral cavity. International Journal of Peptides. 2014;2014:1-13.doi: 10.1155/2014/370297.

16. Vavilova T.P., Yanushevich O.O., Ostrovskaya O.G. Saliva. Analytical possibilities and perspectives. Moscow: BINOM; 2014.

17. Gaspar D, Veiga AS, Castanho MA. From antimicrobial to anticancer peptides. A review. Frontiers in Microbiology. 2013;294(4):1-16. doi:10.3389/fmicb.2013.00294.

18. Dale BA, Fredericks LP. Antimicrobial Peptides in the Oral Environment: Expression and Function in Health and Disease. Current Issues Molecular Biology. 2005;7(2):119-133. doi: 10.1093/jac/dki 103.

19. Budikhina A.S., Pinegin B.V. Defensins are multifunctional human cationic peptides. Immunopathology, allergology, infectology. 2008;2:31-40.

20. Ericksen B, Wu Z, Lu W, Lehrer RI. Antibacterial activity and specificity of the six human alpha-defensins. Antimicrobial Agents and Chemotherapy. 2005;49(1):269-275.doi:10.1128/AAC.49.1.269-275.2005.

21. Abiko Y, Saitoh M, Nishimura M, Yamazaki M, Sawamura D, Kaku T. Role of P-defensins in oral epithelial health and disease. Medical Molecular Morphology. 2007;40(4):79-184. doi: 10.1007/s00795-007-0381-8.

22. Premratanachai P, Joly S, Johnson GK., McCray P, Jia HP, Guthmiller JM. Expression and regulation of novel human defensins in gingival keratinocytes. Oral Microbiology and Immunology. 2004;19(2):111-117. doi: 10.1111/j.0902-0055.2002.00127.x.

23. Lu Q, Jin L, Darveau RP, Samaranayake LP. Expression of human P-defensins-1 and -2 peptides in unresolved chronic periodontitis. Journal of Periodontal Research. 2004;39(4):221-227.doi:10.1111/j. 1600-0765.2004.00727.x.

24. Diamond G, Beckloff N, Weinberg A, Kisich K.O. The roles of antimicrobial peptides in innate host defense. *Current Pharmaceutical Design*. 2009;15(21):2377-2392. doi: 10.2174/138161209788682325.

25. Aleshina G.M., Shamova O.V., Perekrest S.V. et.c Endotoxin-neutralizing effect of antimicrobial peptides // *Cytokines and inflammation*. - 2013. - No. 1 - S. 72-77.

26. Budikhina A.S., Pinegin B.V. Defensins — multifunctional cationic human peptides // *Immunopatol., Allergol., Infectol.* - 2008. - No. 2. — S. 31-40.

27. Lehrer R.I., Lu W. α -Defensins in human innate immunity // *Immunol. Rev.* - 2012. - Vol. 245, No. 1. - P. 84-112.

28. Grishin D.V., Sokolov N.N. Defensins — natural peptide antibiotics of higher eukaryotes // *Biomedical Chemistry*. 2014. V. 60. No. 4. S. 438-447.

29. Li L., Bian T., Lyu J., Cui D., Lei L., Yan F.. Human p-defensin-3 alleviates the progression of atherosclerosis accelerated by *Porphyromonas gingivalis* lipopoly-saccharide // *Int Immunopharmacol*. 2016, Sep. Vol. 38. P. 204-213.

30. Gursoy M., Gursoy U.K., Luukkonen A., Kauko T., Penkkala S., Kononen E. Salivary antimicrobial defensins in pregnancy // *J Clin Periodontol*. 2016 Oct. Vol. 43. No. 10. P. 807-815.

31. Li X., Duan D., Yang J., Wang P., Han B., Zhao L., Jepsen S., Dommisch H., Winter J., Xu Y. The expression of human p-defensins (hBD -1, hBD-2, hBD-3, hBD-4) in gingival epithelia // *Arch Oral Biol*. Jun. 2016 Vol. 66. P. 15-21.

32. Bedi T., Mahendra J., Ambalavanan N. Defensins in periodontal health // *Indian J Dent Res*. 2015. Jul-Aug. Vol. 26. No. 4. P. 340-344.

33. Derradjia A., Alanazi H., Park H.J., Djeribi R., Semlali A., Rouabhia M. α -tocopherol decreases interleukin-1p and -6 and increases human p-defensin-1 and -2 secretion in human gingival fibroblasts stimulated with *Porphyromonas gingivalis* lipopolysaccharide // *J Periodontal Res*. Jun. 2016 Vol. 51. No. 3. P. 295-303.

34. Gursoy U.K., Kononen E., Luukkonen N., Uitto V.J. Human neutrophil defensins and their effect on epithelial cells // *J Periodontol*. 2013. Vol. 84. P. 126-

35. Iskandarovna, Khodzhanova Shakhnoza. "Evaluation Of Platelet Aggregation Activity Depending on The Duration of Antiplatelet Administration in Patients with Coronary Heart Disease." *Journal of Pharmaceutical Negative Results* (2022): 754-760.

36. Khodzhanova Shakhnoza Iskandarovna. (2022). Evaluation Of Platelet Aggregation Activity Depending on The Duration of Antiplatelet Administration in Patients with Coronary Heart Disease. *Journal of Pharmaceutical Negative Results*, 754–760. <https://doi.org/10.47750/pnr.2022.13.S09.086>

37.Кодирова Ш.А., Ходжанова Ш.И. (2022). ФАКТОРЫ РИСКА, ВЛИЯЮЩИЕ НА ТЕЧЕНИЯ ОСТРОГО КОРОНАРНОГО СИНДРОМА. THEORETICAL ASPECTS IN THE FORMATION OF PEDAGOGICAL SCIENCES, 1(6), 109–110. <https://doi.org/10.5281/zenodo.7312294>

38.Rakhmatov A.M., Jabbarov A.A., Kodirova Sh.A., Jumanazarov S.B. (2022). CLINICAL MANIFESTATIONS OF GOUTHY NEPHROPATHY. THEORETICAL ASPECTS IN THE FORMATION OF PEDAGOGICAL SCIENCES, 1(6), 140–141. <https://doi.org/10.5281/zenodo.7322196>

39.Anis, Alyavi, Khodjanova Shakhnoza, and Kadirova Shoir. "Role of the acetylsalicylic acid in the treatment of coronary artery disease." Biomedical Research 31.4 (2020): 82-85.

40.Alyavi, A. L., Khodjanova, S. I., Uzokov, J. K., & Kadirova, S. (2021). Aspirin Resistance in Patients with Chronic Coronary Syndrome. Indian Journal of Forensic Medicine & Toxicology, 15(3), 1843.

41.Ходжанова Ш., Утемуратов Б., Кадырова Ш. АГРЕГАЦИЯ ТРОМБОЦИТОВ И ФАКТОРЫ, ВЛИЯЮЩИЕ НА РЕЗИСТЕНТНОСТЬ К АСПИРИНУ У БОЛЬНЫХ ИШЕМИЧЕСКОЙ БОЛЕЗНЬЮ СЕРДЦА //InterConf. – 2020.

42.Ходжанова Ш. И. ОПРЕДЕЛЕНИЕ АСПИРИНОРЕЗИСТЕНТНОСТИ У БОЛЬНЫХ С ХРОНИЧЕСКИМ КОРОНАРНЫМ СИНДРОМОМ //Боткинские чтения. – 2020. – С. 295-296.

43.Сапаева, З. А., Жабборов, О. О., Максудова, М. Х., Ходжанова, Ш. И., & Кадирова, Ш. А. (2019). Особенности суточного профиля артериального давления у больных системной Красной волчанкой с наличием артериальной гипертензии. Кардиоваскулярная терапия и профилактика, 18(S1), 132-133.

44.Alyavi A. L., Khodjanova S. I. ADP-INDUCED PLATELET AGGREGATION IN PATIENTS WITH CORONARY HEART DISEASE AND WITH ASPIRIN RESISTANCE //Инновационные технологии в медицине: взгляд молодого специалиста. – 2018. – С. 120-121.

45.Ходжанова Ш. И. Оценка функционального состояния сердца и почек у пациентов хронической сердечной недостаточности с дисфункцией почек //Евразийский кардиологический журнал. – 2017. – №. 3. – С. 36-37.

46.Ходжанова Ш. И., Хайитов Х. А., Кодирова Ш. А. Влияние препарата "сулодексид" на функциональное состояние почек у больных хронической болезни почек III стадии на фоне сахарного диабета //Евразийский кардиологический журнал. – 2017. – №. 3. – С. 121-121.

47.Ходжанова Ш. И. и др. КАРДИОРЕНАЛЬНЫЕ ВЗАИМООТНОШЕНИЯ У БОЛЬНЫХ ХРОНИЧЕСКОЙ СЕРДЕЧНОЙ

НЕДОСТАТОЧНОСТЬЮ //О ‘ZBEKISTON TERAPIYA AXBOROTNOMASI.
– С. 23.

48.Кадырова Ш. А., Ходжанова Ш. И. ВЛИЯНИЕ БЛОКАТОРА ГЛИКОПРОТЕИНОВЫХ РЕЦЕПТОРОВ ТРОМБОЦИТОВ П В/ПАТИРОФИБАНА НА КЛИНИЧЕСКОЕ ТЕЧЕНИЕ У БОЛЬНЫХ ОСТРЫМ КОРОНАРНЫМ СИНДРОМОМ БЕЗ ПОДЪЕМА СЕГМЕНТА ST //VIII МЕЖДУНАРОДНОГО КОНГРЕССА «КАРДИОЛОГИЯ НА ПЕРЕКРЕСТКЕ НАУК». – С. 118.

49.Khodjanova S. H. I., Alyavi A. L. International Journal of Pharmacy and Pharmaceutical Science.